

A Novel Deletion in $\delta\beta$ -Thalassemia Found in JapanEiji Matsunaga, Akinori Kimura, Hideo Yamada*
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High molecular weight DNA from a Japanese individual homozygous for $\delta\beta$ -thalassemia was analyzed by the blot hybridization technique of Southern. Results indicated a large deletion of the non- α -globin gene cluster, starting in the vicinity of 3' to the A_γ -globin gene and extending through the 3' side of the β -globin gene. Persistent expression of the γ -globin gene in adult life has been supposed to be caused by loss of a region located about 3-4 kb 5' to the δ -globin gene from comparison of the extents of deletions in several different forms of $\delta\beta$ -thalassemia and HPFH (hereditary persistence of fetal hemoglobin). But the novel deletion found in the present case of $\delta\beta$ -thalassemia suggests that the above putative regulatory region does not have this effect on expression of the γ -globin gene. Some explanations of expression of fetal type globin genes in this $\delta\beta$ -thalassemia are discussed.

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The γ -globin gene is mainly expressed in fetal life and its expression is replaced by those of the δ - and β -globin genes after birth (1). However, in the $\delta\beta$ -thalassemia and HPFH, the γ -globin gene is expressed in adult life (2). In HPFH, synthesis of the γ -globin chain is well balanced by that of the α -globin chain and there are no symptoms characteristic of thalassemia. Expression of the γ -globin gene is insufficient in $\delta\beta$ -thalassemia and the consequent mild imbalance in synthesis of the γ - and the α -globin chains results in slight anemia. Various forms of these two disorders provide good systems for studies on the molecular mechanisms involved in the switch of non- α -globin gene expression ($\gamma \rightarrow \delta, \beta$).

Previous analyses of the gene structure of the non- α -globin gene cluster in the two disorders suggested that the γ -globin gene regulatory region, deletion of which results in persistent expression of the γ -globin gene in adult life (3-5), is located about 3-4 kb 5' to the δ -globin gene, because

this region is absent in all deletion types of HPFH, but not in $\delta\beta$ -thalassemias reported (3-13). This hypothesis on the regulatory region provides an interesting model of the regulation of the globin gene expression during development. To examine this model and elucidate the factors that affect the globin gene expression, we studied a Japanese case of $\delta\beta$ -thalassemia, which was expected to differ from the cases of $\delta\beta$ -thalassemia studied previously in terms of mutation of the non- α -globin gene cluster, because of its different ethnic origin.

Here, we report that this case showed extensive deletion of the non- α -globin gene cluster, including the putative regulatory region.

MATERIALS AND METHODS

Genomic DNA preparation. Total cellular DNAs were isolated from peripheral leukocytes of a Japanese individual homozygous for $\delta\beta$ -thalassemia and from a normal subject (14). The patient with $\delta\beta$ -thalassemia was icteric and showed slight splenomegaly. Hematological data on this patient were as follows: Hb 12.0 g/dl, HbA 0%, HbA₂ 0%, HbF 100% ($G_\gamma/A_\gamma = 0.65/0.35$), with morphological abnormalities of erythrocytes.

Restriction endonuclease mapping of DNA. Total cellular DNAs isolated from the patient and the normal subject were digested completely with several restriction enzymes and then fractionated on 0.6-0.8% agarose gel by electrophoresis. The digests were transferred to nitrocellulose filters and hybridized to appropriate probes labeled with ^{32}P by nick translation (15,16). The probes employed in this study were as follows: $\text{Pst}\beta$ and $\text{Pst}\delta$ are PstI fragments containing the β - and the δ -globin genes, respectively (17). The RIH probe contains a sequence of about 0.5 kb intergenic DNA 4 kb to the 5' side of the δ -globin gene. $\text{Eco}2.3$ is a 2.3 kb EcoRI fragment derived from the 3' flanking region of the A_γ -globin gene (18). $\text{Hin}\gamma$ is a 3.4 kb HindIII fragment including the A_γ -globin gene subcloned from λHyG5 (18). $\text{Bgl}\psi\beta$ is a 4.0 kb BglII fragment containing the $\psi\beta$ -globin gene. The extents of the regions included in these probes are shown in Fig.1.

RESULTS

Deletion of the non- α -globin gene cluster. Genomic DNA from the patient was digested with EcoRI , BamHI and BglII restriction enzymes, fractionated on agarose gels and transferred to nitrocellulose filters (16). The filters were hybridized to $\text{Pst}\beta$, $\text{Pst}\delta$, RIH and $\text{Bgl}\psi\beta$ probes. No specific bands were obtained from the $\delta\beta$ -thalassemia DNA (Fig.2), indicating that the DNA region ranging from the β -globin gene to the $\psi\beta$ -globin gene is entirely deleted on both chromosomes. The deleted region contains the regulatory sequence of the γ -globin gene proposed previously (3-5). The location of this sequence is

A: Normal

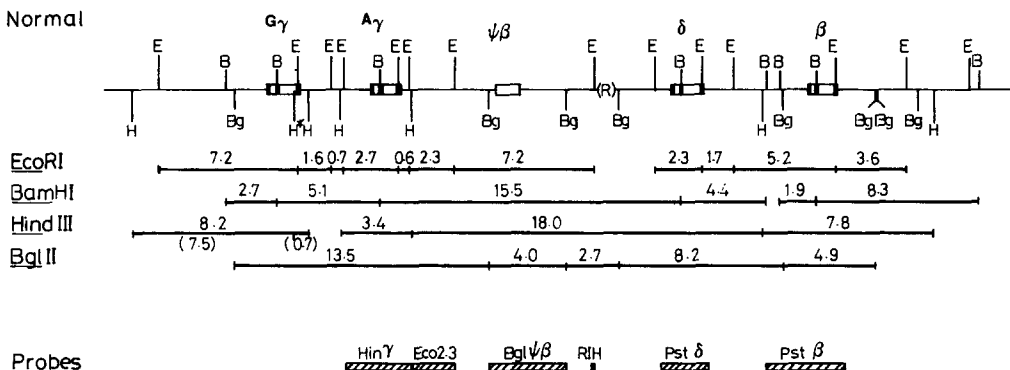
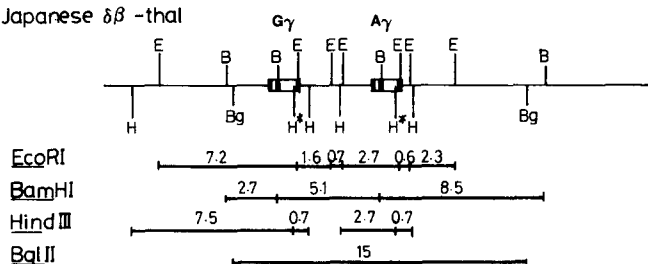
B: Japanese $\delta\beta$ -thal

Fig. 1. Maps of restriction endonuclease cleavage sites within the non- α -globin gene cluster in DNAs of normal subject (A) and the Japanese thalassemia patient (B) with probes used in this study. Abbreviations: H; HindIII, E; EcoRI, B; BamHI, Bg; BglII, H*; HindIII cleavage site polymorphism in the γ -globin genes. (R); the putative regulatory region. The size of the fragments detected by probes are shown in kilobases. The normal map is derived from references 4,6,18 and 19.

shown by (R) in Fig.1. The 3' end point of the deletion could not be defined precisely. However, it is clear that the deletion involved loss of the entire β -globin gene from the fact that the BamHI digest hybridized to the Pst β probe did not generate the 8.3 kb fragment observed with normal DNA (Fig.2).

Extent of the deletion. For determination of the extent of the 5' end point of the deletion in this $\delta\beta$ -thalassemia, the restricted genomic DNA from the patient was hybridized to the HinY probe. EcoRI digests contained 7.2, 2.7, 1.6, 0.7 and 0.6 kb fragments that were identical to those obtained from normal DNA, indicating that the G γ - and the A γ -globin genes are present in the $\delta\beta$ -thalassemia DNA. The faint band of 3.7 kb was formed by cross hybridization of the ϵ -globin gene with the HinY probe. Digests with HindIII hybridized to the same probe gave fragments of 7.5, 2.7 and 0.7 kb instead of

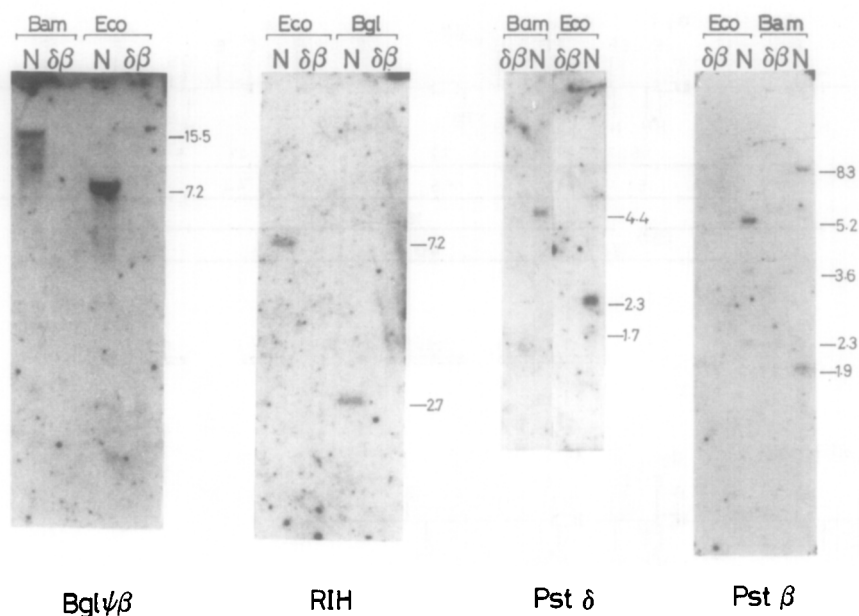


Fig. 2. Hybridizations of DNAs from a normal subject and the Japanese $\delta\beta$ -thalassaemia patient to Pst δ , Pst ϵ , RIH and Bgl $\nu\beta$ probes. The DNA of the patient gave no bands that could be observed with the DNA of the normal subject. N: a normal subject, $\delta\beta$: a Japanese with homozygous $\delta\beta$ -thalassaemia.

the 8.2 and 3.4 kb fragments obtained with normal DNA (Fig.3). This suggests that two HindIII restriction site polymorphisms in both of the γ -globin genes (20) are present on each chromosome in this patient. This observation is consistent with the fact that the entire G_{γ} - and A_{γ} -globin genes are present in this patient and that the patient is homozygous for $\delta\beta$ -thalassemia genotypically. The following abnormal fragments were detected in digests with BamHI and BglII hybridized to the Hinc probe. In BamHI digest, the 15.5 kb fragment found in the normal subject was replaced by a 8.5 kb fragment (Fig.3). This finding is compatible with the extensive deletion of the intergenic region between the A_{γ} - and δ -globin genes (Fig.2), which contains the putative γ -globin gene regulatory region located about 3-4 kb upstream from the δ -globin gene. The BglII digest of thalassemia DNA did not show the normal 13.5 kb fragment containing both the G_{γ} - and A_{γ} -globin genes, but a 15 kb fragment. This observation is consistent with a fact that the BglII fragment containing the $\psi\delta$ -globin gene was deleted from this DNA. Thus the

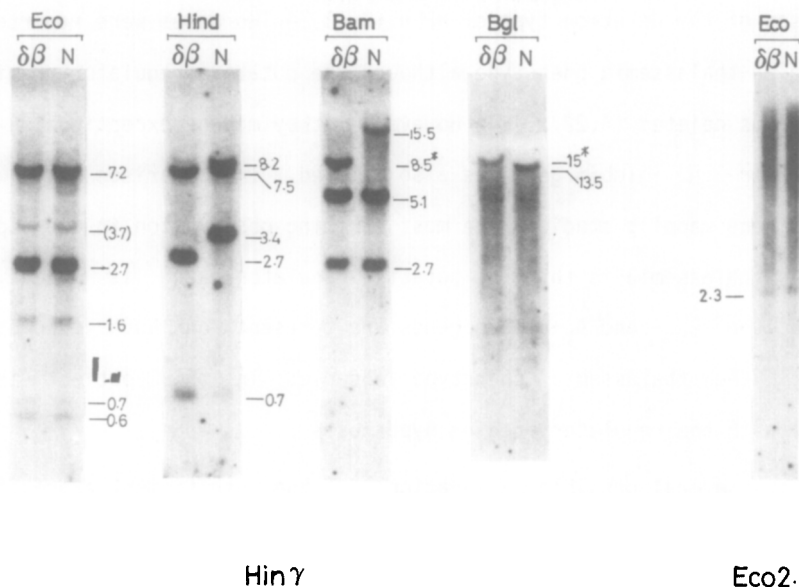


Fig. 3. Hybridizations of normal and Japanese $\delta\beta$ -thalassemia DNAs to $Hin\gamma$ and Eco2.3 probes. The digests with EcoRI and HindIII gave normal bands after hybridization with Eco2.3 and $Hin\gamma$, but the digests with BamHI and BglII hybridized to $Hin\gamma$ gave abnormal bands. The size of the band hybridized with the ϵ -globin gene is given in parenthesis. Asterisks indicate abnormal bands.

BglII site at the 3' end of the normal 13.5 kb fragment is missing in DNA of this $\delta\beta$ -thalassemia (Fig.2). When Eco2.3 was used as a probe, EcoRI digests of the DNAs from the patient and a normal individual, both gave a 2.3 kb band and a smear of hybridization, because Eco 2.3 is known to contain a moderately repetitive sequence (21). These results indicate that the 5' end point of the deletion in this $\delta\beta$ -thalassemia is located in the region between the EcoRI site 2.7 kb 3' to the A_γ -globin gene and the BglII site 0.5 kb 5' to the $\psi\beta$ -globin gene (Fig.1).

DISCUSSION

There are several reports on deletion of the non- α -globin gene cluster in various types of $\delta\beta$ -thalassemia and HPFH (3-13,22,23). It is suggested that the γ -globin gene regulatory region, whose deletion results in persistent expression of the γ -globin gene in adult life, is located between a pair of Alu repeat sequences about 3-4 kb upstream from the δ -globin gene (5), because this region was present in various forms of $\delta\beta$ -thalassemia but absent

in all cases of the deletion type of HPFH (3-13). Two cases were reported to manifest a $\delta\beta$ -thalassemia phenotype although the putative regulatory region in their DNAs was deleted (4,22,23). However, they may be exceptional cases because their A_γ -globin gene was also deleted. In comparison with these results of gene mapping studies, the most striking observation in the Japanese case of $\delta\beta$ -thalassemia is that the putative regulatory area is absent, but that both the G_γ - and A_γ -globin genes are present on both chromosomes. Therefore, the thalassemia phenotype observed in this patient is not consistent with the regulatory region hypothesis.

There are several possible explanations of the insufficient expression of γ -globin gene in this $\delta\beta$ -thalassemia. First, the regulatory sequence might not be present in the region described before, but might be located somewhere beyond the 3' side of the β -globin gene. Tuan *et al.* demonstrated that the 3' end points of the deletion in Sicilian and Turkish cases of $\delta\beta$ -thalassemia were located much closer to the 3' side of the β -globin gene than those of the deletion in HPFH-1 (USA) and HPFH (Ghana) (24). Therefore, some sequence 3' to the β -globin gene present in $\delta\beta$ -thalassemia but not in HPFH could have some role in γ -globin gene expression. Alternatively, in HPFH, the 3' region that was brought into the vicinity of the γ -globin gene by the deletion might contain some sequence enhancing expression of the γ -globin gene (24).

A second possibility is that there is some molecular defect within the γ -globin gene that was not detected by the restriction mapping, but that reduces synthesis of the γ -globin chain.

A third possibility is that one of the two boundaries, which are assumed to form the chromosomal domain essential for expression of the globin genes (9,25), is deleted from the fetal type globin gene domain in this $\delta\beta$ -thalassemia, because of the extensive deletion extending to near A_γ -globin gene.

Some of them could be verified by determination of the nucleotide sequence around the end point of the deletion and the γ -globin gene in DNA of this $\delta\beta$ -thalassemia. Such studies are now in progress. Precise analysis of the

structure of DNA in $\delta\beta$ -thalassemia and HPFH should provide some clues for understanding the mechanism of gene regulation during development.

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REFERENCES

1. Huehns, E.R. (1982) in *Blood and its Disorders*, Hardisty R.M. and Weatherall, D.J. Eds. 2nd ed. pp.323-400, Blackwell Publications, Oxford
2. Weatherall, D.J., and Clegg, J.B. (1981) *The Thalassemia Syndrome*, 3rd ed., Blackwell Scientific Publications, Oxford.
3. Huisman, T.H.J., Schroeder, W.A., Efremov, G.D., Duma, H., Mladenovski, B., Hyman, C.B., Rachmilewitz, E.A., Bouver, N., Miller, A., Brodie, A., Shelton, J.R., Shelton, J.B. and Apell, G. (1974) *Ann. N. Y. Acad. Sci.* 232, 107-124.
4. Fritsch, E.F., Lawn, R.M. and Maniatis, T. (1979) *Nature* 279, 598-603.
5. Ottolenghi, S., Giglioni, B., Taramelli, R., Comi, P., Mazza, U., Saglio, G., Camaschella, C., Izzo, P., Cao, A., Galanello, R., Gimferrer, E., Baiget, M. and Gianni, A.M. (1982) *Proc. Natl. Acad. Sci. USA* 79, 2347-2351.
6. Tuan, D., Biro, P.A., deRiel, J.K., Lazarus, H. and Forget, B.G. (1979) *Nucl. Acids Res.* 6, 2519-2544.
7. Bernards, R., Kooter, J.M. and Flavell, R.A. (1979) *Gene* 6, 265-280.
8. Tuan, D., Murnane, M. J., deRiel, J.K. and Forget, B.G. (1980) *Nature* 285, 335-337.
9. Bernards, R. and Flavell, R.A. (1980) *Nucl. Acids Res.* 8, 1521-1534.
10. Jones, R.W., Old, J.M., Trent, R.J., Clegg, J.B. and Weatherall, D.J. (1981) *Nature* 291, 39-44.
11. Jagadeeswaran, P., Tuan, D., Forget, B.G. and Weissman, S.M. (1982) *Nature* 296, 469-470.
12. Ottolenghi, S. and Giglioni, B. (1982) *Nature* 300, 770-771.
13. Baird, M., Driscoll, M.C., Ben-Bassat, I., Ohta, Y., Nakamura, F., Bloom, A. and Bank, A. (1984) *J. Biol. Chem.* 259, 512-515.
14. Blin, N. and Stafford, D.W. (1976) *Nucl. Acids Res.* 3, 2303-2308.
15. Rigby, P.W.J., Dieckmann, M., Rhodes, C. and Berg, P. (1977) *J. Mol. Biol.* 113, 237-251.
16. Southern, E.M. (1975) *J. Mol. Biol.* 98, 503-517.
17. Lawn, R.M., Fritsch, E.F., Parker, R.C., Blake, G. and Maniatis, T. (1978) *Cell* 15, 1157-1174.
18. Fritsch, E.F., Lawn, R.M. and Maniatis, T. (1980) *Cell* 19, 959-972.
19. Slightom, J.L., Blechl, A.E. and Smithies, O. (1980) *Cell* 21, 627-638.
20. Little, P.F.R., Annison, G., Darling, S., Williamson, R., Camba, L. and Modell, B. (1980) *Nature* 285, 144-147.
21. Fritsch, E.F., Shen, C.K.J., Lawn, R.M. and Maniatis, T. (1980) *Cold Spring Harbor Symposia On Quantitative Biology* 45, 761-775.
22. Jones, R.W., Old, J.M., Trent, R. J., Clegg, J. B. and Weatherall, D. J. (1981) *Nucl. Acids Res.* 9, 6813-6825.
23. Orkin, S.H., Alter, B.P. and Altay, C. (1979) *J. Clin. Invest.* 64, 866-869.
24. Tuan, D., Feingold, E., Newman, M., Weissman, S.M. and Forget, B.G. (1983) *Proc. Natl. Acad. Sci. USA* 80, 6937-6941.
25. Stalder, J., Larsen, A., Engel, J.D., Dolan, M., Groudine, M. and Weintraub, H. (1980) *Cell* 20, 451-460.